



SYNTHESIS AND IN VITRO CYTOTOXICITY OF DIASTEREOMERICALLY MODIFIED DOLASTATIN 15 ANALOGUES

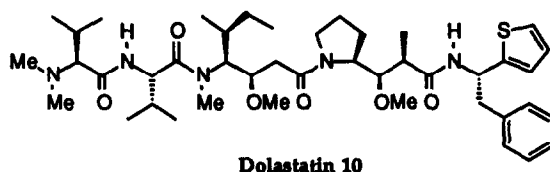
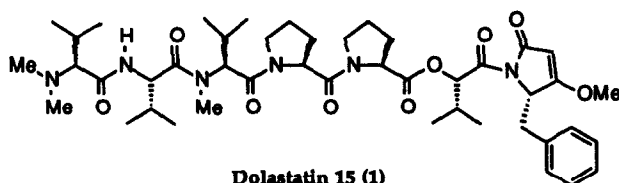
Florence Roux^a, Nathalie Galéotti^a, Joël Poncet^{a*}, Patrick Jouin^a, Suzy Cros^b, Gerhard Zenke^c

^aLaboratoire des Mécanismes Moléculaires des Communications Cellulaires, rue de la Cardonille, 34094 Montpellier Cedex 5, France. ^bLaboratoire de Toxicologie et Pharmacologie Fondamentales, 205 route de Narbonne, 31077 Toulouse Cedex, France. ^cSandoz-Pharma Ltd, CH-4002 Basle, Switzerland.

Key words: Dolastatin; Depsipeptide; Cytostatic activity; Antiproliferative effect

Abstract : Dolastatin 15 **1**(4S,7S) is a potent cytostatic depsipeptide of marine origin. Analysis of the effects of the stereochemistry in the non-peptide moiety on activity revealed that chirality inversion in the aromatic terminal region preserves the antiproliferative activity.

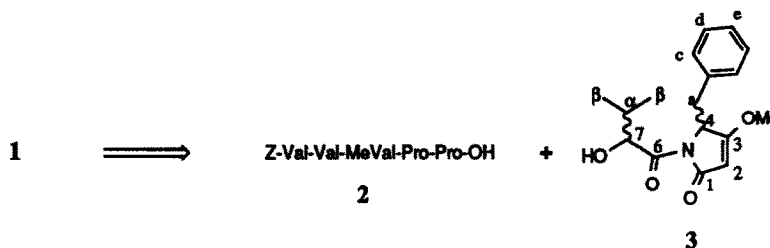
Dolastatin 15 **1**(4S,7S) is a cytostatic depsipeptide isolated from the marine mollusc *Dolabella auricularia*.¹ Its linear structure is related to that of dolastatin 10, another potent antineoplastic metabolite of the same origin.² Detailed biochemical analysis has revealed a qualitatively similar activity profile for both agents, although dolastatin 10 presents higher potency in most studies.³⁻⁵ These agents are antimitotic and specifically interact with tubulin.^{6,7} Their cell cycle phase selectivity was confirmed by typical G2/M metaphase arrest analysis.^{5,8}



The recent development of synthetic procedures for the preparation of dolastatin 10⁹ was accompanied by a preliminary structure-activity analysis.¹⁰ Surprisingly, no such analysis has been performed on dolastatin 15. Having developed an efficient synthesis of dolastatin 15¹¹ we evaluate here the stereoisomeric requirements of the two chiral centers C-4 and C-7¹² present in the non-peptide moiety of the molecule on its cytotoxic activity.

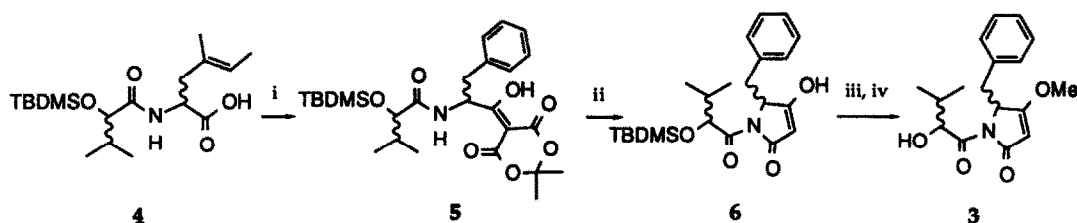
Chemistry

The general convergent strategy used for the preparation of dolastatin 15 (Scheme 1)^{11,13} allows



Scheme 1. Retroynthesis of dolastatin 15 analogues

modifications of acyl-pyrrolidinone 3 and subsequent coupling with the same pentapeptide 2. We obtained the four diastereoisomers of the N-(hydroxy-isovaleryl)-pyrrolidinone 3 by condensation of protected N-(hydroxyisovaleryl)-phenylalanine 4 with Meldrum's acid, cyclisation of the adduct 5 followed by methylation of the corresponding tetramic acid intermediate 6 under Mitsunobu conditions, and finally deprotection of the hydroxyl (Scheme 2).¹¹ Since activation of an N-acylamino acid as in 4 proceeds with epimerisation, we did not attempt to selectively obtain each diastereoisomer of 3 independently. We prepared both 3(4*S*7*S*)/3(4*R*7*S*) and 3(4*S*7*R*)/3(4*R*7*R*) pairs of diastereoisomers starting from N-[(2*S*)-2-hydroxy-isovaleryl]-D,L-phenylalanine 4(7*S*) and the 2*R* counterpart 4(7*R*), respectively. Compound 3(4*S*7*S*) was separated from 3(4*R*7*S*) by silica gel column chromatography. The 3(4*S*7*R*)/3(4*R*7*R*) pair was separated in the same way..



Scheme 2. i: Meldrum's acid (1.1 equiv), dimethylaminopyridine (2.5 equiv), isopropenyl chlorocarbonate (1.1 equiv), CH₂Cl₂, -10 °C, 15 min; ii: Δ, CH₃CN, 30 min, 85% for two steps; iii: Ph₃P (1.2 equiv), MeOH (1.2 equiv), DEAD (1.2 equiv), THF, rt, 16 h, 60%; iv: CF₃CO₂H, rt, 5 min, 85%.

The absolute configuration of compounds 3(4*S*7*S*) and 3(4*R*7*S*) was determined in a previous study.¹¹ The absolute configuration of their respective enantiomers 3(4*R*7*R*) and 3(4*S*7*R*) were assessed by comparison of their physical properties.¹⁴ By using described procedures,¹¹ each alcohol 3 was coupled with pentapeptide 2 and the resulting depsipeptide deprotected and methylated to give the corresponding compound 1.¹⁵

Antiproliferative activity

The *in vitro* antiproliferative potency of both dolastatin 10 and dolastatin 15 has been extensively studied on bone marrow and lymphoma cell lines. Dolastatin 15 has a marked antiproliferative effect on normal human and murine bone marrow progenitor cells and against different human lymphoma cells (IC₅₀, 0.012 to 0.12

nM),^{4,8} as well as on continuous human leukemia cell lines and peripheral blood cells from patients with acute myeloid leukemia (IC₅₀, 0.1 to 1 nM)³. Therefore, we tested the four diastereoisomers of dolastatin 15 for their effect on the proliferation of different human and murine cell lines and primary cells (Table). The assays included growth factor-dependent proliferation of primary murine bone marrow cell, interleukin-2- dependent proliferation of the murine cytotoxic T cell line CTLL, growth factor-independent proliferation of the human leukemia T cell line Jurkat¹⁶ as well as murine leukemia L1210 proliferation¹⁷. As shown in the Table, the compounds inhibit the proliferation of all cell types with IC₅₀ values in the nanomolar range. Dolastatin 15 **1(4S,7S)** was the most potent; the diastereomers **1(4S,7R)** and **1(4R,7S)** were approximately 2-fold less potent and **1(4R,7R)** isomer showed a 10- to 100-fold decreased activity. None of the compounds showed selectivity for any cell type.

Table. Effects of dolastatin 15 (D15) analogues **1** on growth inhibition (IC₅₀, nM)

| | Bone marrow | Jurkat | CTLL | L1210 |
|-----------------|-------------|--------|------|-------|
| 1(4S,7S) | 17 | 5 | 3 | 0.15 |
| 1(4S,7R) | 40 | 10 | 8 | 0.30 |
| 1(4R,7S) | 50 | 10 | 10 | 0.31 |
| 1(4R,7R) | 120 | 700 | 400 | 1.78 |

In the course of an ongoing structure-activity relationship analysis of dolastatin 10 analogues, it was recently demonstrated that inversion of the configuration at the aromatic terminal residue only moderately decreases its cytotoxic activity and its inhibitory effect on tubulin polymerisation.¹⁸ Having prepared dolastatins 10¹⁹ and 15,¹¹ we compared their conformation in solution. We observed a high degree of flexibility in the aromatic region of both compounds. In that context, it is not surprising to observe only a small decrease in activity by inverting the configuration of C-4 in dolastatin 15. Moreover, dolastatin 10 exists in two different conformations corresponding to *cis-trans* isomerisation of the Dil-Dap amide bond (manuscript submitted), whereas all the amide bonds are *trans* in dolastatin 15 (unpublished results). A conformational study of all the analogues presented here is currently under investigation to determine whereas the decrease in growth inhibiting activity could be correlated with a modification of the configuration of one or more of the amide bonds.

Acknowledgement

This research was supported by funds from the Association pour la Recherche sur le Cancer. We are grateful to Dr. S. L. Salhi for help in revising this manuscript.

References and Notes:

1. Pettit, G.R.; Kamano, Y.; Dufresne, C.; Cerny, R.L.; Herald, C.L.; Schmidt, J.M. *J. Org. Chem.* **1989**, *54*, 6005-6006.
2. Pettit, G.R.; Kamano, Y.; Herald, C.L.; Tuinman, A.A.; Boettner, F.E.; Kizu, H.; Schmidt, J.M.; Baczynskyj, L.; Torner, K.B.; Bontems, R.J. *J. Am. Chem. Soc.* **1987**, *109*, 6883-6885.
3. Steube, K.G.; Grunicke, D.; Pietsch, T.; Gignac, S.M.; Pettit, G.R.; Drexler, H.G. *Leukemia* **1992**, *6*, 1048-1053.

4. Jacobsen, S.E.; Ruscetti, F.W.; Longo, D.L.; Keller, J.R. *J. Natl. Cancer Inst.* **1991**, *83*, 1672-1677.
5. Hu, Z.B.; Gignac, S.M.; Quentmeier, H.; Pettit, G.R.; Drexler, H.G. *Leukemia Res.* **1993**, *17*, 333-339.
6. Bai, R.; Pettit, G.R.; Hamel, E. *J. Biol. Chem.* **1990**, *265*, 17141-17149.
7. Bai, R.; Friedman, S.J.; Pettit, G.R.; Hamel, E. *Biochem Pharmacol.* **1992**, *43*, 2637-2645.
8. Beckwith, M.; Urba, W.J.; Longo, D.L. *J. Natl. Cancer Inst.* **1993**, *85*, 483-488.
9. (a) Pettit, G.R.; Singh, S.B.; Hogan, F.; Lloyd, P.; Herald, D.L.; Burkett, D.D.; Clewlow, P. *J. Am. Chem. Soc.* **1989**, *111*, 5463-5465; (b) Tomioka, K.; Kanai, M.; Koga, K. *Tetrahedron Lett.* **1991**, *32*, 2395-2398; (c) Shioiri, T.; Hayashi, K.; Hamada, Y. *Tetrahedron* **1993**, *49*, 1913-1924.
10. Bai, R.; Pettit, G.R.; Hamel, E. *Biochem. Pharmacol.* **1990**, *40*, 1859-1864.
11. Patino, N.; Frérot, E.; Galeotti, N.; Poncet, J.; Dufour, M.-N.; Jouin, P. *Tetrahedron* **1992**, *48*, 4115-4122.
12. Numbering defined in ref. 1.
13. Pettit, G.R.; Herald, D.L.; Singh, S.B.; Thornton, T.J.; Mullaney, J.T. *J. Am. Chem. Soc.* **1991**, *113*, 6692-6693.

14. Physical and spectral data of 3.

| Compound | [α] _D ²⁰ (c 1, MeOH) | H ¹ NMR, 360 MHz (DMSO <i>d</i> ₆) δ (ppm) | | | | | | | |
|------------------|--|--|------------|------------|------|-----------|------|------|-----------|
| | | H β | H α | H α | OMe | H6 | H4 | H2 | Hc,d,e |
| 3(4S7S)[3(4R7R)] | +313(-300) | 0.82, 0.88 | 1.85-1.96 | 3.00, 3.50 | 3.82 | 4.82-4.84 | 4.90 | 5.08 | 6.92-7.25 |
| 3(4R7S)[3(4S7R)] | -232(+244) | 0.85, 0.95 | 1.82-1.90 | 3.10, 3.30 | 3.84 | 4.76-4.77 | 4.90 | 5.14 | 6.92-7.24 |

15. Physical and spectral data of 1 (H¹ NMR data are consistent with the assigned structures).

| Compound | [α] _D ^a | mp (°C) | HPLC, rt (mn) ^b | MS(FAB+) m/e ^c |
|----------|--|-----------|----------------------------|---------------------------|
| 1(4R7S) | -285 | 159 - 161 | 5.50 | 838 (9) |
| 1(4R7R) | -290 | 151 - 153 | 6.66 | 838 (19) |
| 1(4S7R) | -25 | 140 - 142 | 6.75 | 838 (9) |

a: c 0.2, MeOH; b: C8, Ultrabase 5 μ m, B(CH₃CN, 0.1% TFA)/A(H₂O, 0.1% TFA), 1.5 ml/mn, 40 to 70% B in 10 mn; c: MH⁺(relative intensity).

16. Schreier, M.H.; Zenke, G.; Borel, J.F.; Baumann, G. *Transplant. Proc.* **1992**, *24*, 19-21.

17. L1210 leukemia cells were grown in nutrient medium RPMI 1640 (GIBCO) supplemented with 20% heat inactivated horse serum, L-glutamine (2 mM), penicillin (200 U/ml), and streptomycin (50 μ g/ml). Cultures were incubated in a 5% CO₂-air incubator at 37 °C. Various concentrations of the drugs were prepared in ethanol (0.5% final concentration), and 0.02 ml of each solution was added to 1.98 ml of leukemic cells in log phase, at an initial density of 0.8 10⁵ cells/ml. Control groups were tested in four wells; each drug concentration was tested in triplicate. Cell proliferation was checked using a Coultronics Coulter Counter. Cell counting was performed after 48 hours of culture. The results were analysed by a regression curve of the probit of the percent of growth inhibition versus the log concentration used. The concentration causing 50% growth inhibition (IC₅₀) was determined. All experiments were repeated three times.

18. Bai, R.; Roach, M.C.; Jayaram, S.K.; Barkoczy, J.; Pettit, G.R.; Luduena, R.F.; Hamel, E. *Biochem. Pharmacol.* **1993**, *45*, 1503-1515.

19. Roux, F.; Maugras, I.; Poncet, J.; Niel, G.; Jouin, P. *Tetrahedron* **1994**, *50*, 5345-5360.

(Received in Belgium 25 March 1994; accepted 20 June 1994)